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TERRECYCLIC ACID A, A NEW ANTIBIOTIC FROM ASPERGILLUS TERREUS

III. ¹⁸C NMR SPECTRUM OF TERRECYCLIC ACID A

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Assignment of the fifteen carbons of terrecyclic acid A, $C_{15}H_{20}O_8$, a new sesquiterpene antibiotic, in the ¹⁸C NMR spectrum was performed by ¹⁸C-{¹H} selective proton decoupling experiments, comparison with spectra of its derivatives and chemical shifts.

Terrecyclic acid A (I), $C_{15}H_{20}O_3$, is a sesquiterpene antibiotic from *Aspergillus terreus* Thom No. 14^{1,2)} and has the same novel carbon skeleton as does quadrone, an antitumor substance from *A. terreus*^{3,4)} (Fig. 1). *A. terreus* No. 14 also produces several substances related to I^{5,6)}. We are much interested in the biosynthesis of I and of related compounds.

Assignments of all carbons of I in ¹³C NMR spectrum are necessary not only for studying biosynthesis but also for characterizing related compounds which will be isolated in the future.

The ¹⁸C NMR spectral data for I are shown in Table 1. Since there are two signals in the region of the carbonyl carbons, chemical shifts readily assigned the C-7 and 4 carbons at δ 179.95 and 207.54, respectively. Also from the chemical shift the singlet at δ 150.53 must be due to C-5. The remaining

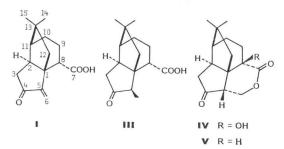
Carbon	I	III	IV	V
1	54.90 (s)	54.93 (s)	54.49 (s)	49.87 (s)
2	46.36 (d)	45.02 (d) ^b	47.39 (d)°	45.98 (d) ^d
3	41.45 (t)	40.45 (t)	43.00 (t)	43.20 (t)
4	207.54 (s)	218.83 (s)	218.10 (s)	216.57 (s)
5	150.53 (s)	51.60 (d) ^b	52.13 (d)°	52.51 (d) ^d
6	116.10 (t)	8.57 (q)	66.98 (t)	65.29 (t)
7	179.95 (s)	181.24 (s)	172.49 (s)	174.01 (s)
8	47.94 (d)	48.03 (d) ^b	78.36 (s)	48.73 (d) ^d
9	22.52 (t)	22.17 (t)	28.72 (t) ^a	19.33 (t)
10	28.87 (t)	28.84 (t)	29.89 (t) ^a	28.08 (t)
11	48.85 (d)	49.32 (d) ^b	50.81 (d)°	52.21 (d) ^d
12	54.03 (t)	47.68 (t)	46.71 (t)	52.51 (t)
13	40.45 (s)	39.55 (s)	40.34 (s)	40.45 (s)
14	27.32 (q)	27.17 (q)	26.82 (q)	26.91 (q)
15	34.75 (q)	34.19 (q)	34.84 (q)	34.84 (q)

Table 1. Summary of ¹³C NMR spectral data of I, III, IV and V (25 MHz, CDCl₃) (δ, ppm, multiplicity).

^a Assignments may be reversed.

b, c, d Assignments may be changed.

Fig. 1. Structures of I, III, IV and V.



two quaternary carbons (C-1 and 13) were assigned mainly from the LINDEMAN-ADAMS rule⁷⁾. According to this rule C-1 and 13 should resonate at δ 47.09 and 38.29, respectively. The difference between the calculated and found values comes from the β -deshielding effect, because the LINDEMAN-ADAMS rule applies to paraffin molecular structure. Consequently, C-1 appears at δ 54.90, while C-13 appears at δ 40.45.

There are three methine carbons and assignments of them could be made by ¹⁸C-{¹H} selective proton decoupling experiments. The 100 MHz ¹H NMR spectrum of I has already

	Hn Hg Hg Hc Hn Hg Hc Hn Hg Hc OOH	
Proton	Chemical shift (õ, ppm, multiplicity)	Coupling constant (Hz)
=CH ₂	{5.96 (1H, s) (5.21 (1H, s)	
H_{f}	3.01 (1H, d)	J = 8.0
H _c	2.95 (1H, dd)	J=10.0, 11.0
H_{d} *	2.65 (1H, dd)	J=11.0, 19.5
H_{e}^{*}	2.53 (1H, dd)	J=10.0, 19.5
H_{g}	2.14 (1H, m)	
H_h	1.97 (1H, dd)	J = 2.8, 2.8
H_k	1.90 (1H, m)	
H_j and H_1	1.78~1.85 (2H, m)	
$H_{\rm m}$ and $H_{\rm n}$	1.77 (2H, s)	
^{н₃с} ×	{1.24 (3H, s) 1.18 (3H, s)	

Table 2. Summary of ¹H NMR spectrum of I.

* Assignment may be reversed.

been reported, but the resolution was not complete¹⁾. That was improved in the 400 MHz ¹H NMR spectrum (Fig. 2). Each proton was labeled according to that in the diazomethane adduct of the methyl ester of I (II), the ¹H NMR spectrum of which was completely analyzed²⁾. Assignment of the protons necessary for ¹³C-{¹H} selective proton decoupling experiments was made as follows*.

By comparison of the spectra of I and II, the signals at δ 3.01 (1H, d) and at δ 1.97 (1H, dd) are attributable to the H_f and H_h protons, respectively. Since the H_f proton is affected by the carboxyl group, it is deshielded in the ¹H NMR spectrum. From coupling constants the H_e, H_d and H_e protons are assigned as shown in Table 2, but the assignment of H_d and H_e protons may be reversed.

Using the results of these assignments ¹³C-{¹H} selective proton decoupling experiments were conducted. On irradiation at $\delta_{\rm H}$ 2.90 the signal at $\delta_{\rm c}$ 46.36 was sharpened, and on the other hand, irradiation at $\delta_{\rm H}$ 3.00 sharpened the $\delta_{\rm c}$ 47.94 resonance. Thus the $\delta_{\rm c}$ 46.36 and 47.94 resonances are due to C-2 and 8, respectively. The remaining methine carbon, C-11, was assigned as the resonance at δ 48.85.

There exist five methylene carbons (C-3, 6, 9, 10 and 12) in I and the δ 116.10 resonance can readily be attributed to C-6, an olefinic carbon. Assignment of C-3 was made by ¹⁸C-{¹H} selective proton

^{*} Assignment of other protons was made as follows. Two singlets at δ 5.96 and 5.21 are readily assigned to the exomethylene protons. Judging from the characteristic shape of signals in the spectra of I and II the multiplet at δ 2.14 is assigned to the H_g proton and the multiplet at δ 1.90 to the H_k proton. In the ¹H NMR spectrum of II the H_m and H_n protons show typical AB type doublets (*J*=15.5 Hz), but in that of I these two protons are equivalent and appear as a singlet at δ 1.77. On the other hand the two methyl groups are non-equivalent, giving a 3H-singlet at δ 1.24 and a 3H-singlet at δ 1.18, while in II they are equivalent. Accordingly, the remaining 2H overlapped multiplet at δ 1.78~1.85 is assigned to the H_J and H₁ protons. The results are summarized in Table 2.

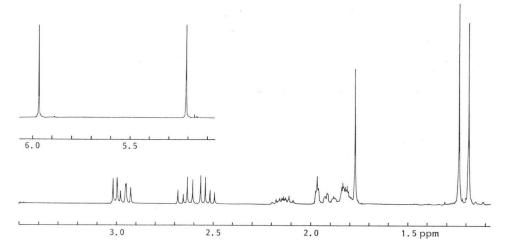


Fig. 2. 400 MHz ¹H NMR spectrum of I (CDCl₃, TMS).

decoupling experiment. Since on irradiation of $\delta_{\rm H}$ 2.60 the signal at $\delta_{\rm c}$ 41.45 was sharpened, the C-3 carbon was assigned as δ 41.45.

By catalytic hydrogenation of I over palladium-charcoal, the reduced product (III), $C_{15}H_{22}O_3$, was obtained in high yield⁵⁾. The relative stereochemistry of I has already been determined⁵⁾, and the new methyl group of III should be on the same side as the C-12 methylene carbon, because catalytic hydrogenation occurs on the side of less steric hindrance.

The ¹³C NMR spectral data for III are shown in Table 1. There are four methylene carbons (C-3, 9, 10 and 12) and comparison of the chemical shifts of the methylene carbons in ¹⁸C NMR spectra of I and III indicates the carbon signals at δ 22.52, 28.87, 41.45 and 54.03 in I shift upfield to δ 22.17, 28.84, 40.45 and 47.68 in III, respectively. Since C-12 in III is most affected by the steric compression effect of a new methyl group (C-6), C-12 appears at δ 47.68 in III and appears at δ 54.03 in I.

Furthermore, we have isolated some compounds related to I from the ethyl acetate neutral fraction of the culture filtrate of *A. terreus* No. 14 and have elucidated their structures⁶). One of them is 8-hydroxyquadrone (IV), whose ¹³C NMR spectrum is shown in Table 1. There also exist five methylene carbons (C-3, 6, 9, 10 and 12) in quadrone (V) and IV. It is presumed by comparison to the ¹³C NMR spectrum of I that the remaining two methylene carbons (C-9 and 10) resonate at δ 19.31 and 28.08 in V and at δ 28.72 and 29.89 in IV. In 8-hydroxyquadrone the hydroxyl group shifts the C-9 resonance about 8 ppm downfield by its β -deshielding effect. Accordingly, C-9 resonates at δ 19.31 in V and at δ 22.52 in I.

Chemical shifts of the two methyl carbons in I are δ 27.32 and 34.75. According to the Dreiding stereomodels, C-14 exists spatially near C-9, 10, 12 and 15, and on the other hand C-15 exists near C-3, 12 and 14. Judging from the steric compression effect the C-14 resonance can be assigned as δ 27.32.

Assignments of all carbons in the 13 C NMR spectrum of terrecyclic acid A are shown in Table 1, and the carbons of **III**, **IV** and **V** are also assigned, but assignments of the latter three compounds are in some cases tentative.

Terrecyclic acid A seems to be a sesquiterpene antibiotic with an abnormal carbon skeleton and now we are studying the biosynthesis of I on the basis of the data reported in this paper.

Experimental

Melting points were determined on a microscope hot stage of Yanagimoto Co. and are uncorrected. The optical rotation was measured with a Jasco DIP-SL polarimeter. The IR spectra were recorded on a Jasco IRA-2 infrared spectrophotometer. The ¹H NMR spectra were obtained with Jeol JNM-FX-400 and FX-100 spectrometers. The ¹³C NMR spectra were measured with a Jeol JNM-FX-100 spectrometer. The mass spectra and high resolution mass spectrum were obtained with a Hitachi RMU-6M and a Jeol JMS D-300 spectrometer, respectively.

Reduction Product from I (III)

Reaction conditions have been reported in detail⁵). Physico-chemical properties of **III** are as follows: mp 128 ~ 129°C; $[\alpha]_D^{28} - 27^\circ$ (*c* 0.4, EtOH); MS *m*/*z* (relative abundance) 250 (M⁺, 8.1), 235 (9.5), 232 (2.0), 193 (100), 163 (10.7), 121 (10.1); IR ν_{max}^{KBr} cm⁻¹ 3150 (br), 2940, 1710 (br), 1450, 1380, 1290, 1240, 1190, 1160, 1060, 1040, 855.

8-Hydroxyquadrone (IV)

The isolation procedure will be reported in detail⁶). Physico-chemical properties of IV are as follows: mp 136~137°C; $[\alpha]_D^{34}$ -51° (*c* 0.3, EtOH); MS *m*/*z* 264.1378 (M⁺, 264.1361 calcd for C₁₅H₂₀O₄), 248 (M-16), 236 (M-28), 220 (M-44); IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹ 3450, 1740, 1730.

Quadrone (V)

Quadrone was obtained by pyrolysis of I^{D} : mp 183~184°C, $[\alpha]_{\text{D}}^{\text{D}}$ -44.6° (c 1.3, EtOH).

Addendum in Proof

Quite recently the study on the biosynthesis of terrecyclic acid A using the results of this paper has been reported by the present authors⁵⁾.

Acknowledgments

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